#### real radiofrequency pulses

Up to now the effects of rf pulses are described with neglection of further interactions (mainly chemical shift evolution) during the pulses. In this case the spin operators are rotated around the spin operator given by the pulse phase in the rotating frame.

In the following section we will analyze the more real situation where chemical shift evolution takes place during the action of pulses.

For a one spin system, any arbitrary spin state can be expressed by

$$\sigma(t) = c_x(t) \boldsymbol{I}_x + c_y(t) \boldsymbol{I}_y + c_z(t) \boldsymbol{I}_z$$

This state can be described by the corresponding coefficient vector:

$$\sigma(t) = \begin{bmatrix} \boldsymbol{I}_{x} \boldsymbol{I}_{y} \boldsymbol{I}_{z} \end{bmatrix} \begin{bmatrix} c_{x}(t) \\ c_{y}(t) \\ c_{z}(t) \end{bmatrix}$$

To calculate the time development of the system it is sufficient to analyze the evolution of this coefficient vector.

The action of pulses with x- or y-phases and the larmor precession results in rotations of the coefficient vector around the corresponding axes. The rotation angle is given by  $\phi = \omega \tau$ 

These rotations can be represented as matrices acting on the coefficient vector:

$$R_{x} = \begin{bmatrix} 1 & 0 & 0 \\ 0 & \cos \phi & -\sin \phi \\ 0 & \sin \phi & \cos \phi \end{bmatrix} = \begin{bmatrix} 1 & 0 & 0 \\ 0 & \cos \omega \tau & -\sin \omega \tau \\ 0 & \sin \omega \tau & \cos \omega \tau \end{bmatrix}$$

$$R_{y} = \begin{bmatrix} \cos \phi & 0 & \sin \phi \\ 0 & 1 & 0 \\ -\sin \phi & 0 & \cos \phi \end{bmatrix}$$

$$R_z = \begin{bmatrix} \cos \phi & -\sin \phi & 0 \\ \sin \phi & \cos \phi & 0 \\ 0 & 0 & 1 \end{bmatrix}$$

The evolution of the coefficient is then given by

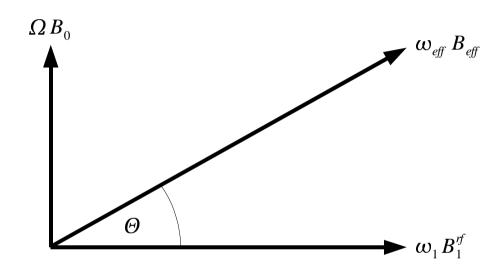
$$\begin{bmatrix} c_{x}(\tau) \\ c_{y}(\tau) \\ c_{z}(\tau) \end{bmatrix} = R_{\alpha}(\omega, \tau) \begin{bmatrix} c_{x}(0) \\ c_{y}(0) \\ c_{z}(0) \end{bmatrix}$$

Are two fields, e.g. rf-field and  $B_0$ -field, active at the same time, an effective field results with direction and strength given by simple vector addition:

$$\omega_{eff} B_{eff} = \omega_1 B_1^{rf} + \Omega B_0$$

$$\omega_{eff} = \sqrt{\omega_1^2 + \Omega^2}$$

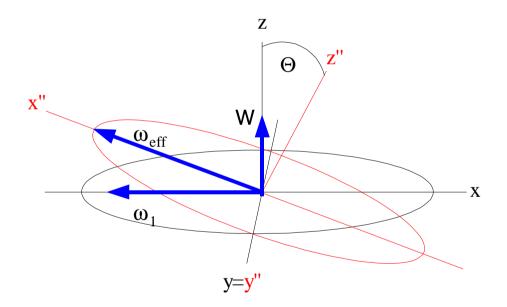
$$\cos \Theta = \frac{\omega_1}{\omega_{eff}}$$



The magnetization precesses now around the effective field  $B_{\it eff}$  with a frequency  ${\bf w}_{\it eff}$  .

The precession around the effective field can be described by three subsequent rotations:

- 1. rotation around y (assumption: x-pulse) with an angle  $-\Theta$ ,
- 2. rotation around x" with an angle  $\phi = \omega_{eff} \tau$
- 3. rotation around y = y'' with an angle  $\Theta$



$$\begin{bmatrix} c_x(\tau) \\ c_y(\tau) \\ c_z(\tau) \end{bmatrix} = R_y(\Theta) R_x(\phi) R_y(-\Theta) \begin{bmatrix} c_x(0) \\ c_y(0) \\ c_z(0) \end{bmatrix} = \begin{bmatrix} \cos \Theta & 0 & \sin \Theta \\ 0 & 1 & 0 \\ -\sin \Theta & 0 & \cos \Theta \end{bmatrix} \begin{bmatrix} 1 & 0 & 0 \\ 0 & \cos \phi & -\sin \phi \\ 0 & \sin \phi & \cos \phi \end{bmatrix} \begin{bmatrix} \cos \Theta & 0 & -\sin \Theta \\ 0 & 1 & 0 \\ \sin \Theta & 0 & \cos \Theta \end{bmatrix} \begin{bmatrix} c_x(0) \\ c_y(0) \\ c_z(0) \end{bmatrix}$$

multiplying alk three rotation matrices result in the general rotation matrix describing the action of an arbitrary rectangular pulse with x-phase with simultanuous presence of larmor precession around the z-axis:

$$R_{P}(\Omega, \omega_{1}, \tau) = \begin{bmatrix} \cos^{2}\Theta + \cos\phi \sin^{2}\Theta & -\sin\phi \sin\Theta & -\cos\Theta \sin\phi (1 - \cos\phi) \\ \sin\phi \sin\Theta & \cos\phi & -\sin\phi \cos\Theta \\ \cos\Theta \sin\phi (1 - \cos\phi) & \sin\phi \cos\Theta & \sin^{2}\Theta + \cos\phi \cos^{2}\Theta \end{bmatrix}$$

In the case of an arbitrary phase  $\alpha$  of the rf-field in the x,y plane an additional rotation around the z-axis is necessary:

$$R_P(\Omega, \omega_{1,} \tau, \alpha) = R_z(\alpha) R_P(\Omega, \omega_{1,} \tau) R_z(-\alpha)$$

With these rotation matrices it is possible to calculate the excitation profile of any given pulse.

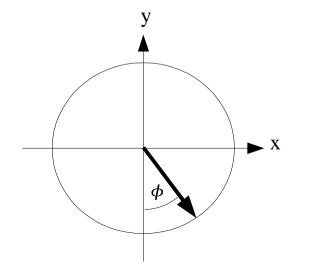
For the action of rf-pulses three rotations are important:

1. Excitation,

rotating of z-magnetization into the transverse plane, this is the standard on-resonant 90° pulse

$$I_{z} \xrightarrow{\omega_{1} \tau I_{x} + \Omega_{I} \tau I_{z}} c_{x}(\tau) I_{x} + c_{y}(\tau) I_{y} + c_{z}(\tau) I_{z}$$
with  $\omega_{1} \tau = \frac{\pi}{2}$  for  $\Omega_{I} = 0$  result in 
$$\begin{bmatrix} c_{x}(\tau) \\ c_{y}(\tau) \\ c_{z}(\tau) \end{bmatrix} = \begin{bmatrix} 0 \\ -\sin(\omega_{1}\tau) \\ \cos(\omega_{1}\tau) \end{bmatrix} = \begin{bmatrix} 0 \\ -1 \\ 0 \end{bmatrix}$$

Instead of viewing the cartesian der x,y,z components of the magnetization the excitation profile of an excitation pulse can be decribed by the amplitude and phase of the resulting transverse magnetization:



$$M_{transversal} = M_0 \sqrt{c_x^2(\tau, \Omega_I) + c_y^2(\tau, \Omega_I)}$$

$$\tan(\phi, \Omega_I) = -\frac{c_x(\tau, \Omega_I)}{c_y(\tau, \Omega_I)}$$

2. inversion rotation from z nach -z (or opposite direction) by a 180° pulse with x- or y-phase

3. refocusing rotation from x to -x by a 180° pulse with y-phase, or rotation from y nach -y by a 180° pulse with x-phase

The influence of larmor precession during the action of the pulse is not the same in the case of inversion compared to refocusing. Consequently a 180° pulse of same duration shows different profiles for both rotations.

For the inversion the pulse phase does not play a role, because the resulting transverse magnetization comes from pulse imperfections and should be suppressed by proper methods-

A refocusing pulse acts on transverse magnetization, therefore the phase of the pulse is important.

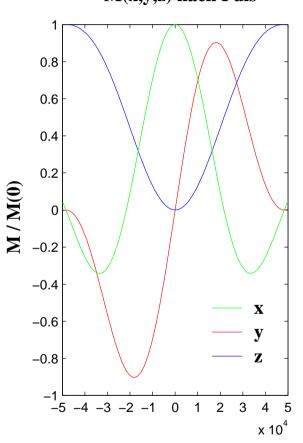
The following rule is valid:

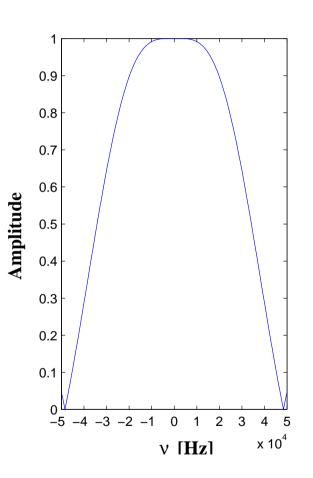
a good refocusing pulse is also a good inversion pulse, but a good inversion pulse might be a worse refocusing pulse

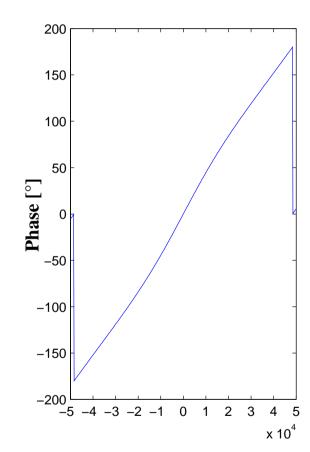
### Excitationprofile of a 90° rectangular pulse

pulse duration 20 µs 
$$v_{rf} = \frac{\gamma B_1}{2\pi} = \frac{1}{4\tau_{90}} = 12500 Hz$$

#### M(x,y,z) nach Puls







Excitation in the region  $|\nu| < \nu_{rf} \rightarrow \sqrt{M_x^2 + M_y^2}$ 

Excitation in the region  $|v| < 1.5 v_{rf} \rightarrow \sqrt{M_x^2 + M_y^2} / M_0 > 0.91$ 

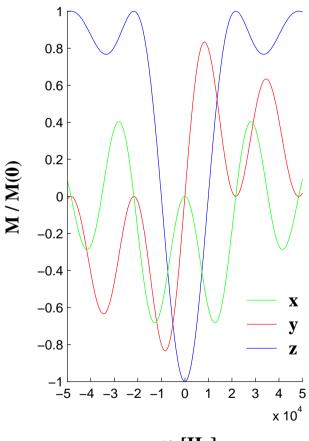
1. excitation 'zero' ( $M_{x,y} = 0$ ) at  $v_{zero} \approx \frac{0.97}{\tau_{90}}$ 

#### Inversionprofile of a 180° rectangula pulse

pulse duration 40 µs 
$$\rightarrow v_{rf} = \frac{\gamma B_1}{2\pi} = \frac{1}{4\tau_{90}} = 12500 Hz$$

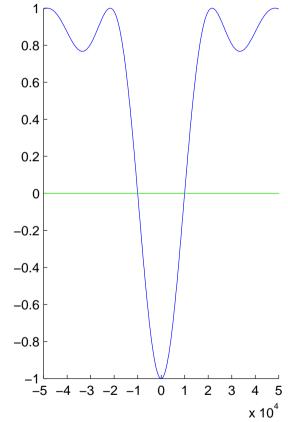
M(x,y,z) nach Puls

M(x,y,z) nach Phasenzyklus 1. Experiment:  $\phi = +x$ , rec = + 2. Experiment:  $\phi = -x$ , rec = +



ν **[Hz]** 

inversion in the region  $|v| < 0.25 v_{rf} : M_z \rightarrow -M_z > 0.90$ inversion in the region  $|v| < 0.5 v_{rf} : M_z \rightarrow -M_z > 0.55$ 



1. inversion 'zero' 
$$(M_z = 0)$$
 at  $v_{zero} \approx \frac{0.87}{\tau_{180}}$ 

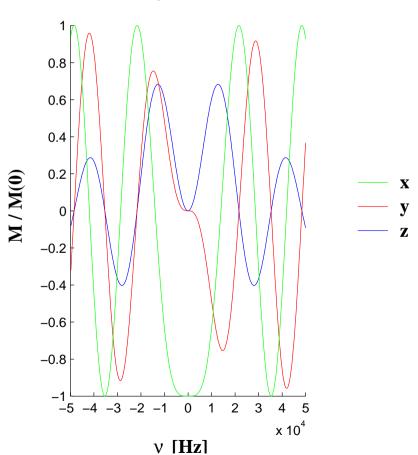
# 

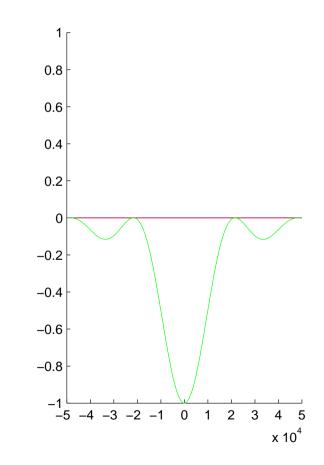
pulse duration 40 µs 
$$\rightarrow v_{rf} = \frac{\gamma B_1}{2\pi} = \frac{1}{4\tau_{90}} = 12500 Hz$$

#### M(x,y,z) nach Phasenzyklus

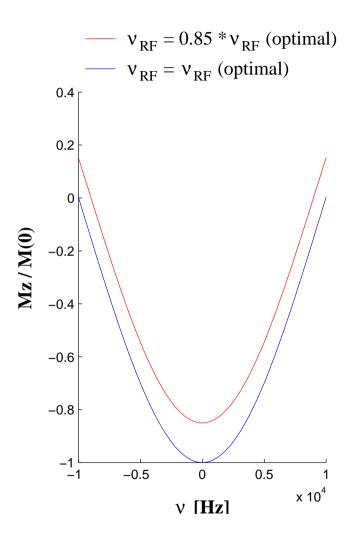
- 1. Experiment:  $\phi = +x$ , rec = +
- 2. Experiment:  $\phi = -x$ , rec = +
- 2. Experiment:  $\phi = y$ , rec = -
- 2. Experiment:  $\phi = -y$ , rec = -

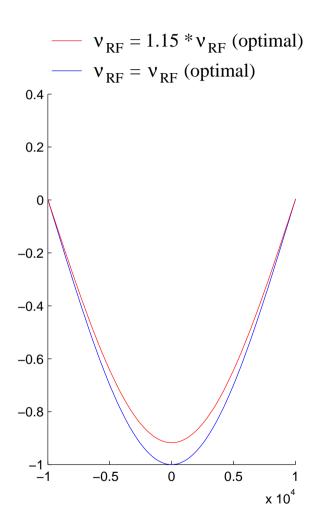
M(x,y,z) nach Puls





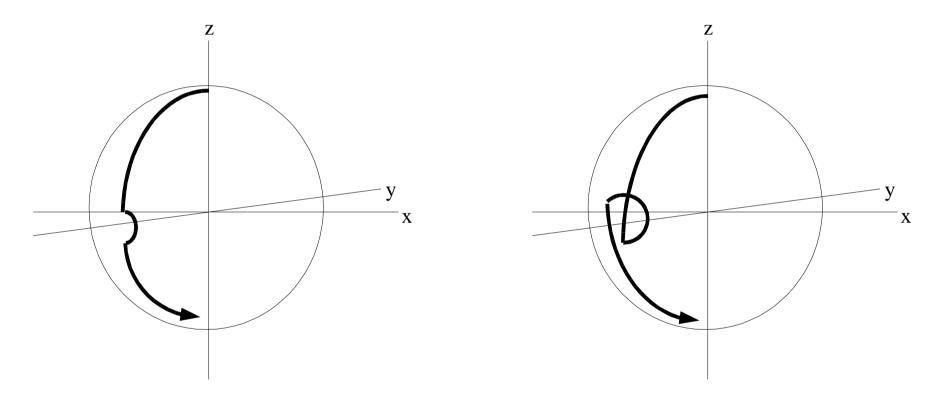
# bad calibrated 180° pulse





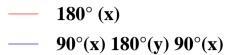
pulse length too short

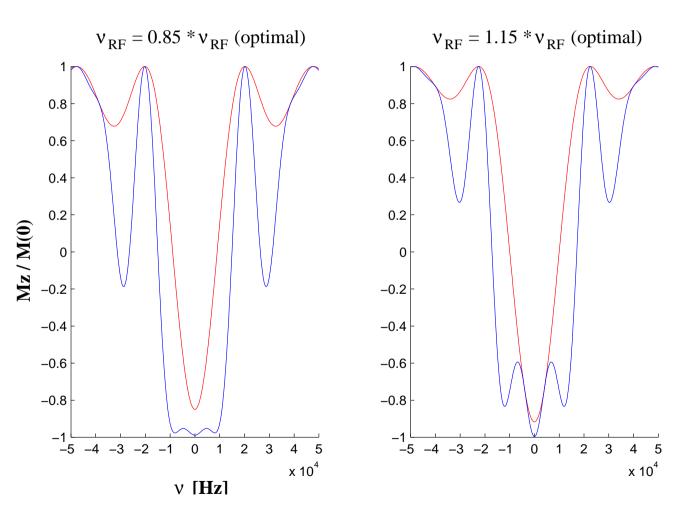
pulse length too long



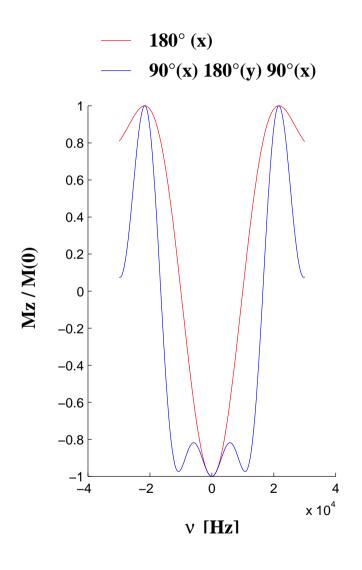
composite pulse compensates the bad calibration to a large extent

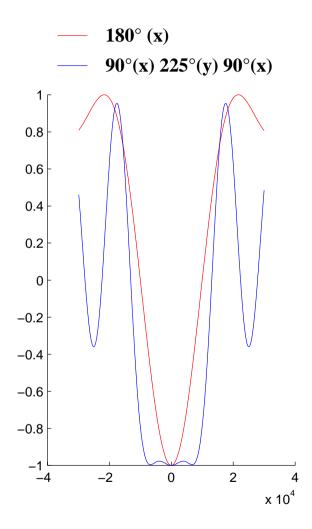
Inversion profile of a 'bad' calibrated 90°(x) 180°(y) 90°(x) composite pulse





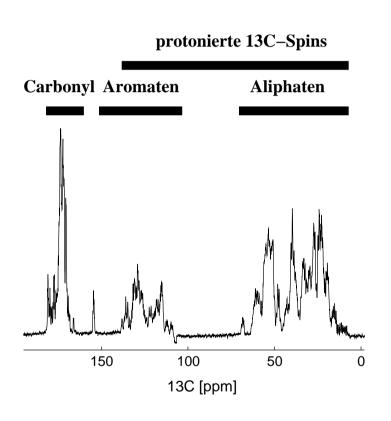
# Inversion profile of a correct calibrated composite pulse

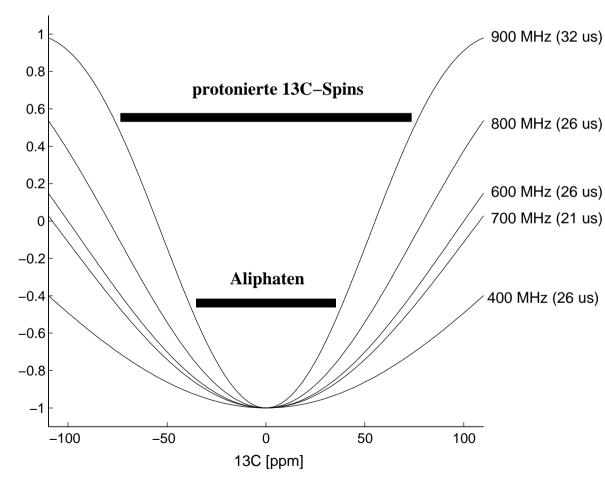




# <sup>13</sup>C NMR on proteins and real pulses

#### Inversions profil eines high–power 13C 180 $^{\circ}$ Pulses





The well separated regions of <sup>13</sup>C' and <sup>13</sup>C<sup>a</sup> chemical shifts in proteins allow the selective rotations by proper selected pulses.

A prominent method is to choose the durations of 90° and 180° pulses that they only excite only one of both regions, while the excitation profile of the applied pulse shows an excitation minimum in the other frequency range.

A 90° pulse has a first excitation minimum at ca. 0.97 / t, a  $180^\circ$  pulse (inversion) has a first minimum at ca. 0.87 / t

The center of the <sup>13</sup>C' resonances lies around 175 ppm, that of the <sup>13</sup>C<sup>a</sup> spins around 56 ppm.

For a 600 MHz spectrometer the following pulse lengths should be used:

$$\tau_{90} = \frac{0.97}{119 \times 151 \, Hz} = 54 \, \mu \, s$$
$$\tau_{180} = \frac{0.87}{119 \times 151 \, Hz} = 48 \, \mu \, s$$

Remark: a 108 ms long 180° pulse has a too narrow inversion profile to be used for protein triple resonance experiments.

#### Amplitude- and phasemodulated pulses

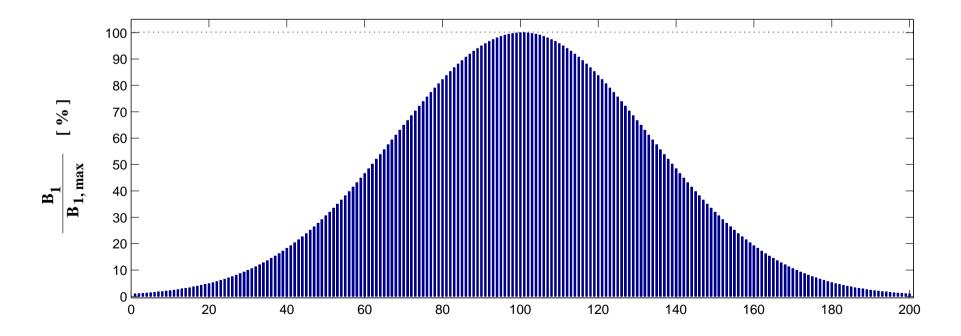
modern spectrometer allow not only using rectangular pulses, that means a constant amplitude during the pulse, but also pulses with variable amplitude and phase during the pulse can be realized.

With these 'shaped' pulses very designed excitation profiles can be achieved.

These pulses are realized by a seried of very short rectangular pulses wih different amplitudes and phases.

The time resolution of each pulse steps is around 50 ns on modern spectrometers.

Example: Gauß shape like puls, consisting of 200 short rectangular pulses



#### Calculation of the exciation profile of an amplitude modulated pulse

For a pulse consisting of N short rectangular pulses with equal length and variable amplitude  $A_i$  (in the range of  $0.0 - 1.0 * \omega_1(max)$ ), the excitation profile can be calculated (numerically) by subsequent application of the corresponding rotation matrices for the individual short rectangular pulses acting on the actual states of the magnetization:

$$\boldsymbol{\tau}_{p} = N \cdot \Delta \boldsymbol{\tau}$$

$$M(\boldsymbol{\tau}_{p}) = \left\{ \prod_{i=1}^{N} R(\boldsymbol{\omega}_{1}^{i}, \Delta t, \boldsymbol{\phi}_{i}, \Omega) \right\} M(0)$$

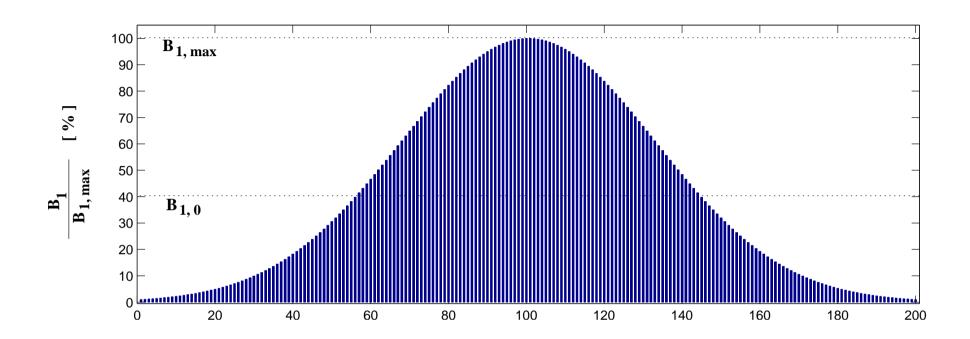
The onresonant pulse angle in the case of an amplitude modulated pulse (phase =  $180^{\circ}$  corresponds to a negative amplitude) is given by

$$\alpha = \sum_{i=1}^{N} \alpha_i$$
 mit  $\alpha_i = \omega_1^i \Delta \tau$ 

Using the integral ratio (= ratio between the overall amplitude of a rectangular pulse with equal length and  $\omega_1^0$  and the integrated amplitude of the shaped pulse) and the onresonant pulse angle  $\alpha$  the necessary rf field strength  $\omega_1^{max}$  can be calculated:

$$Int.Ratio = \frac{1}{\sum_{i=1}^{N} A_{i}} \qquad \omega_{1}^{0} = \frac{\alpha}{\tau_{p}} \qquad \longrightarrow \qquad \omega_{1}^{max} = \frac{\omega_{1}^{0}}{Int.Ratio} \qquad \omega_{i} = A_{i} \omega_{1}^{max}$$

# Example: Gauß-like 90° pulse



*Int.Ratio* = 0.40997

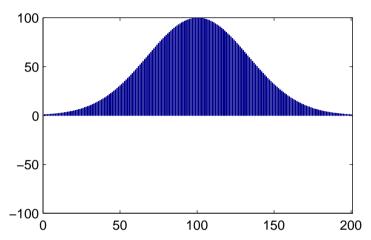
$$\tau_p = 512 \mu s$$

$$\frac{\omega_1^0}{2\pi} = 488.3 \, Hz$$

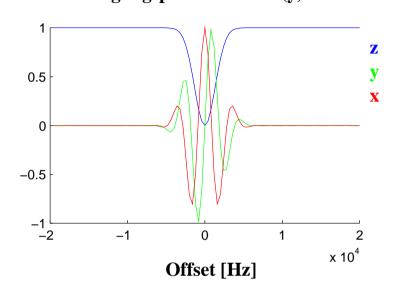
$$\frac{\omega_1^{max}}{2\pi} = 1191.0 \, Hz \quad \text{(corresponds to a } 210 \, \mu \, s \, 90^{\circ} \, \text{rectangular pulse)}$$

# Excitation profile of the Gauss pulse (90°, 512 ms)

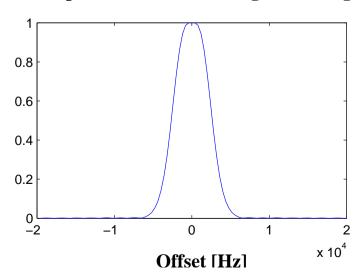




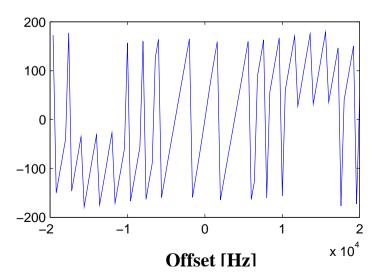
#### Anregungsprofil nach 90°(y) Puls

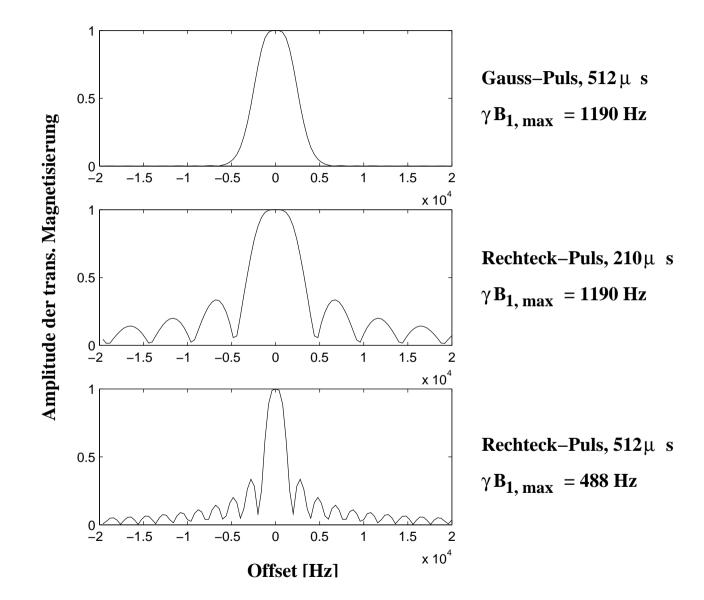


# Amplitude der transv. Magnetisierung



#### Phase der transvers. Magnetisierung





# Summary: Comparison of a gauss-shape pulse with a rectangular pulse

In contrast to the rectangular pulse the Gauss-shape pulse exhibits the following features

At the same  $B_1^{max}$  the Gauss-like pulse exhibits a more narrow excitation profile The Gauss-shape pulse shows up no further exciation bands outside the central band.

The rectangular pulse is better for exciting a broader frequency band at a given rf field strength

The Gauss-shape pulse is better for a more selective excitation band at a given rf field strength

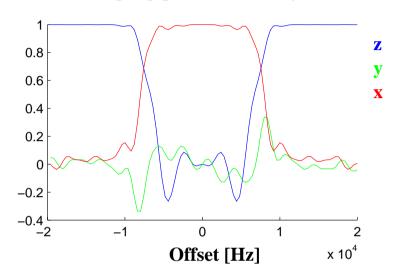
# Excitation profile of a 90° G4 pulse (Gaussian cascade), 512 ms

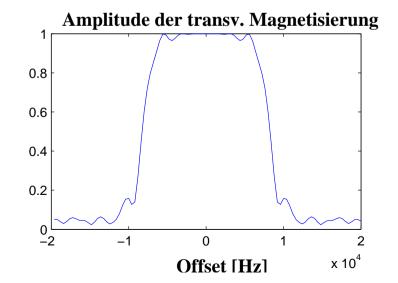
→ nearly top hat like excitation

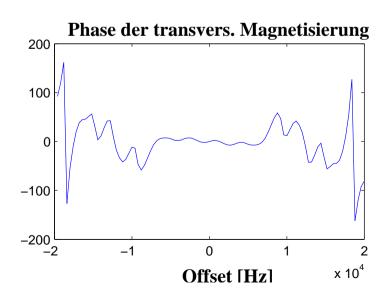


# 100 50 -50 -100 0 50 100 150 200

#### Anregungsprofil nach 90°(y) Puls

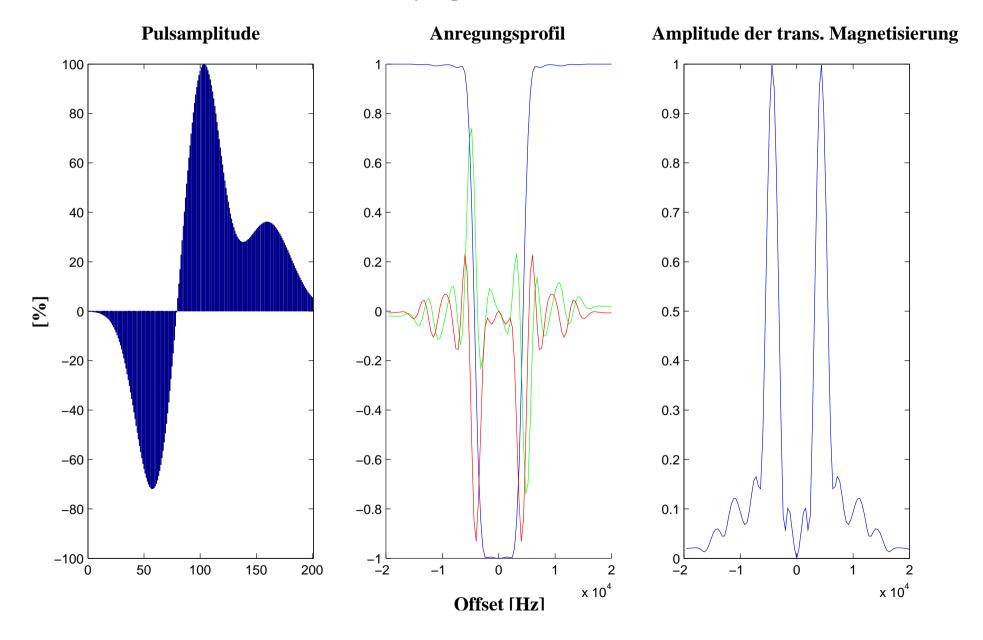






# Inversion profile of 180° G3 pulse (Gaussian cascade, 512 ms)

→ nearly top hat like inversion



#### Coherence order and coherence transfer selection

Coherences in NMR (transversal magnetization) can be classified by their coherence order p, this is the number of  $I^+$  operators (positive coherence), and  $I^-$  operators (negative coherence)

example: 
$$I_x = \frac{1}{2}(I^+ + I^-)$$
  $\longrightarrow p = 1$  und  $p = -1$ 

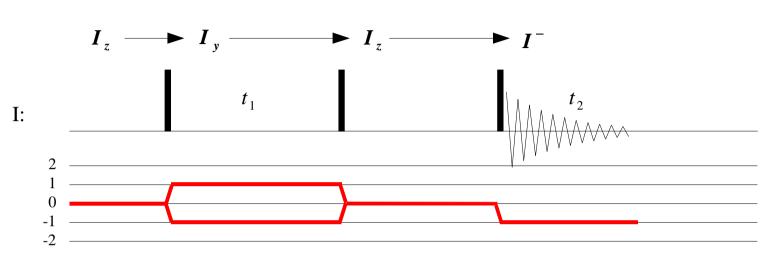
$$I_z \qquad \longrightarrow p = 0$$

$$2I_x S_x = \frac{1}{4}(I^+ + I^-)(S^+ + S^-) \qquad \longrightarrow p = 2$$
 und  $p = 0$  und  $p = -2$ 

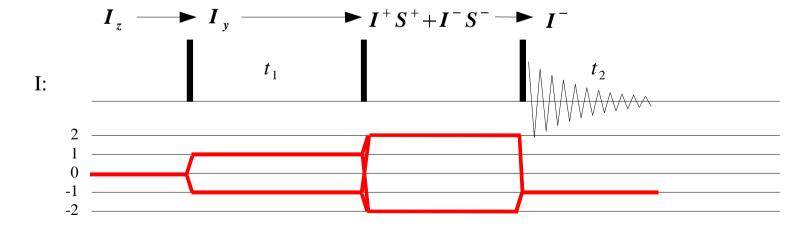
$$= \frac{1}{4}[I^+ S^+ + I^+ S^- + I^- S^+ + I^- S^-]$$

Goal in a (multidimensional) NMR experiment is properly selecting a defined coherence transfer pathway and suppressing all other transfer pathway in order to get an easy interpretable spectrum.

#### A) NOESY

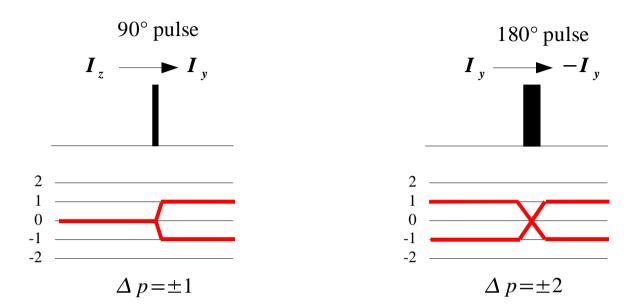


## B) DQF-COSY



→ How can the desired coherence transfer pathway be selected?

#### 1. Pulses change the coherence order



2. Changing the r.f. phase (pulse phase) alters phase of the coherence

$$I_z \xrightarrow{\frac{\pi}{2}I_y} I_x = \frac{1}{2}[I^+ + I^-]$$

$$\Delta \phi_{rf} = \frac{\pi}{2} \qquad \boldsymbol{I}_{z} \xrightarrow{-\frac{\pi}{2} \boldsymbol{I}_{x}} \boldsymbol{I}_{y} = -\frac{i}{2} [\boldsymbol{I}^{+} + \boldsymbol{I}^{-}] = \frac{1}{2} [\exp\{-i\frac{\pi}{2}\}\boldsymbol{I}^{+} + \exp\{i\frac{\pi}{2}\}\boldsymbol{I}^{-}]$$

general: a phase alteration of a pulse by  $\Delta \phi_{rf}$  results in a phase shift of the coherence with order p by  $\Delta \phi = -p \Delta \phi_{rf}$ 

By systematic variation of the rf-phases together with the receiver phase (phase cycling) in subsequent single experiments it is possible to select desired magnetization transfer pathways and suppressing the unwanted transfers by difference spectroscopy.

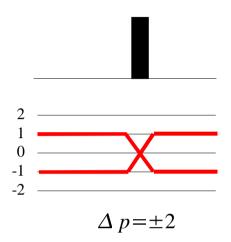
Example: phase cycling in the DQF-COSY

the first both pulses shall create  $Dp = \pm 2$  (generation of double quantum coherence)

	$oldsymbol{\phi}_{\it vf}$	$\phi_{_{\Deltap=+2}}$	$\phi_{\Delta p=-2}$	$oldsymbol{\phi}_{rec}$
X	$0^{\circ}$	$0^{\circ} = 0^{\circ}$	$0_{\circ} = 0_{\circ}$	0°
у	90°	$-180^{\circ} = 180^{\circ}$	$180^{\circ} = 180^{\circ}$	180°
-X	180°	$-360^{\circ} = 0^{\circ}$	$360^{\circ} = 0^{\circ}$	$0^{\circ}$
<b>-y</b>	270°	$-540^{\circ} = 180^{\circ}$	$540^{\circ} = 180^{\circ}$	180°

Behaviour of unwanted transfers in this phase cycle:

Example: Elimination of imperfections of a 180° refocusing pulse (EXORCYCLE)



$$I^+ \longrightarrow I^+(\Delta p=0)+I^-(\Delta p=-2)+I_z(\Delta p=-1)$$

$$I^- \longrightarrow I^-(\Delta p=0)+I^+(\Delta p=2)+I_z(\Delta p=1)$$

	$oldsymbol{\phi}_{r\!f}$	$\phi_{_{\Deltap=+2}}$	$\phi_{_{\Deltap=-2}}$	$oldsymbol{\phi}_{rec}$
X	$0^{\circ}$	$0_{\circ} = 0_{\circ}$	$0_{\circ} = 0_{\circ}$	0°
y	90°	$-180^{\circ} = 180^{\circ}$	$180^{\circ} = 180^{\circ}$	180°
-X	180°	$-360^{\circ} = 0^{\circ}$	$360^{\circ} = 0^{\circ}$	$0^{\circ}$
<b>-y</b>	270°	$-540^{\circ} = 180^{\circ}$	$540^{\circ} = 180^{\circ}$	180°

Behaviour of unwanted transfers in this phase cycle:

#### General aspects of phase cycles

The minimal phase cycle determines the necessary number of scans per  $t_1$ -increment

With many pulses it is no more practicable to alter all rf-phases independently, because the number of required accumulations will increase the overall experimental time.

It is not nice if the necessary number of accumulations for phase cycle is larger than required for a given sample concentration. In this case the experimental time is longer than necessary for generating sufficient intensity.

Phase cycles act as difference methods. If a very intense signal must be eliminated by phase cycling, instabilities cause a non perfect suppression of these signal, because of small signal intensity variations in subsequent scans.

#### Pulsed field gradients

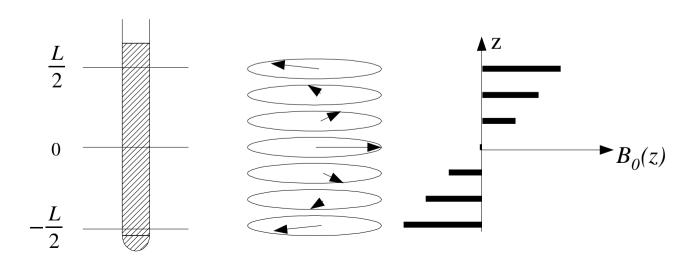
pulsed field gradients are short (pulse duration: 0.1-2 ms) defined variations of the external static  $B_0$  field, causing defined spatial inhomogenities during these gradient pulses.

$$B_0(z) = B_0 + z \frac{\Delta B}{\Delta z}$$

- resonance frequency depends on spatial position:  $\omega_0(z) = -\gamma B_0(z) = -\gamma B_0 \gamma z G_z$
- After the gradient pulse the coherences have aquired a spatial depending phase:

$$I^{+} \xrightarrow{\omega_{0}(z)\tau_{grd}I_{z}} I^{+} \exp\{izG_{z}\tau_{grd}\} = I^{+} \exp\{i\phi_{grd}(z)\} \quad \text{with } \phi_{grd}(z) = yzG_{z}\tau_{grd}$$

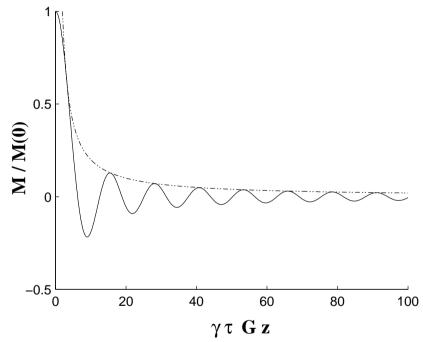
$$I^{-} \xrightarrow{\omega_{0}(z)\tau_{grd}I_{z}} I^{-} \exp\{-izG_{z}\tau_{grd}\} = I^{-} \exp\{-i\phi_{grd}(z)\}$$



The macroscopic observable signal is given by the integrated signal intensity over the sample volume:

$$I^{+}(\tau_{grd}) = \frac{1}{L} I^{+}(0) \int_{-\frac{L}{2}}^{\frac{L}{2}} \exp\{i \gamma G_{z} \tau_{grd} z\} dz$$

$$= I^{+}(0) \frac{\sin(\gamma G_{z} \tau_{grd} L/2)}{\gamma G_{z} \tau_{grd} L/2} = I^{+}(0) \operatorname{sinc}(\gamma G_{z} \tau_{grd} L/2)$$



For sufficient long gradient pulses the decay of macroscopic observed coherencedue to dephasing by the gradient is given by  $2/(\gamma G_z L \tau_{grd})$ 

With a sample length of 4 cm and a typical gradient strength of 10 Gauss / cm the macroscopic observable coherence is reduced to less than 0.2 % with a 1 ms gradient pulse .

transverse magnetization can be dephased very efficiently by pulsed field gradients.

Behaviour of mixed product operators under action of pulsed field gradients

$$I^{+}S_{z}$$
  $\xrightarrow{\gamma_{I}zG_{z}\tau_{grd}I_{z}}$   $I^{+}S_{z}\exp\{i\gamma_{I}zG_{z}\tau_{grd}\}=I^{+}S_{z}\exp\{i\phi_{grd}^{I}(z)\}$ 

$$I^{+}S^{+} \xrightarrow{y_{I}zG_{z}\tau_{grd}I_{z}} \xrightarrow{y_{S}zG_{z}\tau_{grd}S_{z}} I^{+}S^{+} \exp\{i\gamma_{I}zG_{z}\tau_{grd}\}\exp\{i\gamma_{S}zG_{z}\tau_{grd}\}$$

$$= I^{+}S^{+}\exp\{i(\phi_{grd}^{I}(z)+\phi_{grd}^{S}(z))\}$$

$$I^{+}S^{-} \xrightarrow{\gamma_{I}zG_{z}\tau_{grd}I_{z}} \xrightarrow{\gamma_{S}zG_{z}\tau_{grd}S_{z}} I^{+}S^{-}\exp\{i\gamma_{I}zG_{z}\tau_{grd}\}\exp\{-i\gamma_{S}zG_{z}\tau_{grd}\}\}$$

$$=I^{+}S^{-}\exp\{i(\phi_{grd}^{I}(z)-\phi_{grd}^{S}(z))\}$$

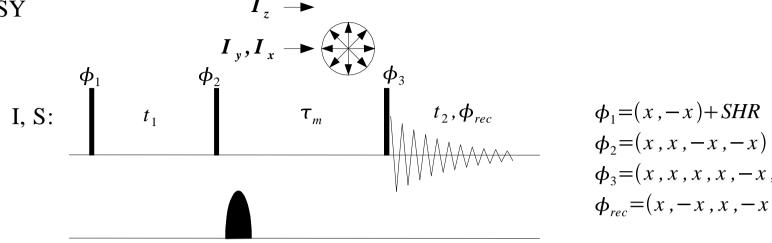
The resulting spatial dependent phase depends linear on the coherence order

$$\phi_{grd}(z) = p \gamma z G_z \tau_{grd}$$

pulsed field gradients allow the discrimination of different coherences within a single scan!!

Dephasing of undesired transverse magnetization with pulsed field gradients

**NOESY** 



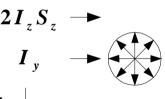
$$\phi_{1} = (x, -x) + SHR$$

$$\phi_{2} = (x, x, -x, -x)$$

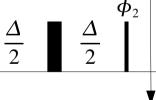
$$\phi_{3} = (x, x, x, x, -x, -x, -x, -x)$$

$$\phi_{rec} = (x, -x, x, -x, -x, -x, x, x, -x)$$

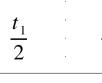
**HSQC** 

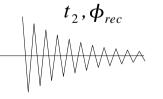


S:



 $\mathbf{v} \boldsymbol{\phi}_1$ 





decoupling

$$\Delta \approx \frac{1}{2J_{IS}}$$

$$\phi_1 = \{x, -x\} + SHR$$

$$\phi_2 = \{ y, y, -y, -y \}$$

$$\phi_{rec} = \{x, -x, -x, x\}$$

#### Defined de- and rephasing with pulsed field gradients

If a desired coherence (transverse magnetization) is dephased by a pulsed field gradien, a second gradient must rephase the spatial variation along the sample in order to get an observable signal:

example:

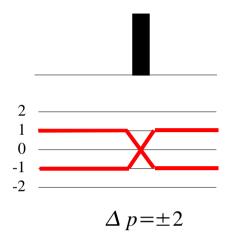
$$I^+ \xrightarrow{G_A(z)} I^+ \exp\{i\phi_A(z)\} \longrightarrow I^+ \exp\{i\phi_A(z)\} \xrightarrow{G_B(z)} I^+ \exp\{i\phi_A(z)\} \exp\{i\phi_B(z)\}$$

to achieve a macroscopic observable signal, the second gradient must compensate the dephasing caused by the first gradeint:

$$\exp\{i\phi_A(z)\}\exp\{i\phi_B(z)\} = \exp\{i(\phi_A(z) + \phi_B(z))\} = 1 \quad \text{that means } \phi_A(z) + \phi_B(z) = 0$$

By setting strength and duration of different gradients defined selections of coherence transfer pathways within a single scan are possible

#### Selection of refocusing within a single scan by a pulsed field gradient



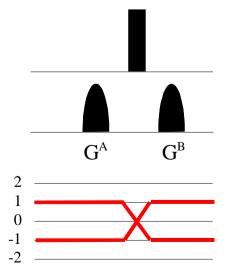
$$I^+ \longrightarrow I^+(\Delta p=0)+I^-(\Delta p=-2)+I_z(\Delta p=-1)$$

$$I^- \longrightarrow I^-(\Delta p=0)+I^+(\Delta p=2)+I_z(\Delta p=1)$$

Suppression of undesired transfers by phase cycling 

→ EXORCYCLE

four independent scans necessary

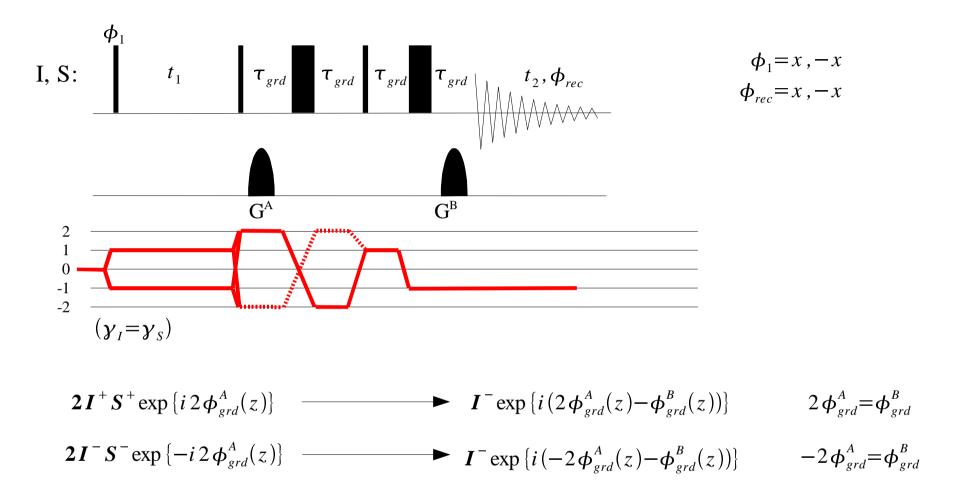


$$I^{+} \longrightarrow I^{+} \exp\left\{i\left(\phi_{grd}^{A} + \phi_{grd}^{B}\right)\right\} + I^{-} \exp\left\{i\left(\phi_{grd}^{A} - \phi_{grd}^{B}\right)\right\} + I_{z} \exp\left\{i\left(\phi_{grd}^{A}\right)\right\}$$

$$I^{-} \longrightarrow I^{-} \exp\left\{-i\left(\phi_{grd}^{A} + \phi_{grd}^{B}\right)\right\} + I^{+} \exp\left\{-i\left(\phi_{grd}^{A} - \phi_{grd}^{B}\right)\right\} + I_{z} \exp\left\{-i\left(\phi_{grd}^{A}\right)\right\}$$

 $G^A = G^B$  selects the refocusing within a single scan and suppresses the unwanted transfers

#### Coherence selection in the DQF-COSY by pulsed field gradients



 $\longrightarrow$  Only the transfer  $2I^+S^+ \rightarrow I^-$  or the transfer  $2I^+S^+ \rightarrow I^-$  can be selected in a single scan, but not both!

The spinecho  $\tau_{grd}$  - 180° -  $\tau_{grd}$  serves for refocusing of the evolution of chemical shifts during the duration of the gradient (avoids a large first order phase error)

#### Coherence selection in the HSQC by pulsed field gradients

I: 
$$\frac{\Delta}{2} \begin{bmatrix} \frac{\Delta}{2} \end{bmatrix} \begin{bmatrix} t_2, \phi_{rec} \\ \phi_1 = \{x, -x\} \end{bmatrix} \\ \phi_1 = \{x, -x\} \end{bmatrix}$$

$$S: \begin{bmatrix} \frac{t_1}{2} & \frac{t_1}{2} & \tau_{grd} \end{bmatrix} \begin{bmatrix} \tau_{grd}$$

 $+\frac{1}{2}[(-\frac{1}{2})\mathbf{I}^{-}]\exp\{i(\Omega_{S}t_{1}-\phi_{1}(z))\} + \frac{1}{2}[(-\frac{1}{2})\mathbf{I}^{-}]\exp\{i\Omega_{S}t_{1}\}\exp\{i\Omega_{I}t_{2}\}\exp\{-i(\phi_{1}(z)+\phi_{2}(z))\}$ 

in order to obtain a macroscopic observable signal the spatial dependent phases caused by the two gradients must be cancel each other:

$$\phi_1(z) = \phi_2(z)$$
 $\Rightarrow$  selection of transfer

 $I_z S^+ \rightarrow \frac{1}{4} I^ Y_S \tau_{grd} G_1 z = y_I \tau_{grd} G_2 z$ 

$$\frac{G_1}{G_2} = \frac{\gamma_I}{\gamma_S}$$

under the assumption that the duration of both gradients are equal

or 
$$-\phi_1(z) = \phi_2(z)$$
  $\longrightarrow$  selection of transfers  $I_z S^- \to \frac{1}{4} I^-$  
$$-\gamma_S \tau_{grd} G_1 z = \gamma_I \tau_{grd} G_2 z$$

 $-\frac{G_1}{G_2} = \frac{\gamma_I}{\gamma_S}$  under the assumption that the duration of both gradients are equal

In both cases a phasemodulated signal results: 
$$Signal(t_1,t_2) = \frac{1}{4} \exp\left\{-i\Omega_S t_1\right\} \exp\left\{i\Omega_I t_2\right\} \ \ \text{mit} \quad \boldsymbol{\gamma}_S G_1 = \boldsymbol{\gamma}_I G_2$$
 
$$Signal(t_1,t_2) = \frac{1}{4} \exp\left\{i\Omega_S t_1\right\} \exp\left\{i\Omega_I t_2\right\} \ \ \text{mit} \quad \boldsymbol{\gamma}_S G_1 = -\boldsymbol{\gamma}_I G_2$$

With the alternation of the polarity (sign) of G1 (or G2) for a given  $t_1$  inkrement a phaseble hypercomplex signal can be created by linear combination:

$$Signal_{N}(t_{1}, t_{2}) = \frac{1}{4} \exp \left\{-i \Omega_{S} t_{1}\right\} \exp \left\{i \Omega_{I} t_{2}\right\} \quad \text{mit} \quad \boldsymbol{\gamma}_{S} G_{1} = \boldsymbol{\gamma}_{I} G_{2}$$

$$Signal_{P}(t_{1}, t_{2}) = \frac{1}{4} \exp \left\{i \Omega_{S} t_{1}\right\} \exp \left\{i \Omega_{I} t_{2}\right\} \quad \text{mit} \quad \boldsymbol{\gamma}_{S} G_{1} = -\boldsymbol{\gamma}_{I} G_{2}$$

$$Signal_{\cos}(t_{1}, t_{2}) = Signal_{P}(t_{1}, t_{2}) + Signal_{N}(t_{1}, t_{2}) = \frac{1}{2} \cos \left(\Omega_{S} t_{1}\right) \exp \left\{i \Omega_{I} t_{2}\right\}$$

$$Signal_{\sin}(t_{1}, t_{2}) = i \left(Signal_{P}(t_{1}, t_{2}) - Signal_{N}(t_{1}, t_{2})\right) = \frac{1}{2} \sin \left(\Omega_{S} t_{1}\right) \exp \left\{i \Omega_{I} t_{2}\right\}$$

$$\text{,,echo-antiecho transformation''}$$

After the echo-antiecho transformation the resulting twodimensional FID shows the same signal intensity like a 'classical' HSQC, recorded phase sensitive by the SHR method,

#### but

during the echo-antiecho transformation the noise from both single experiments adds up, and for the noise level results::

$$|Noise_{\cos, \sin}| = |Noise_P + Noise_N| = \sqrt{2} |Noise_P|$$
  $\longrightarrow$   $S/N(SHR) = \sqrt{2} S/N(grd. selection)$ 

Comparing the two coherence selections with gradients in the experiment HSQC and DQF COSY the following aspects result:

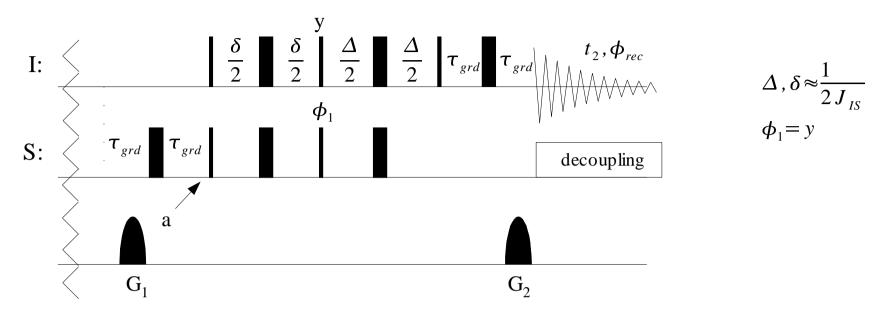
- 1. Selection by gradients during a fixed time interval (e.g. i the DQF-COSY) reduces the signal intensity by a factor of 2 compared to the selection with a proper phase cycling, because only the positive or the negative coherence results in observable signal.
- 2. Selection by a gradient during the evolution period reduces the overall signal intensity to  $\frac{1}{\sqrt{2}}$  signal (phase cycling), if only one of both cartesian components of the desired coherence is transferred to the final observed signal.

The question now is, whether the transfer

$$I_z S^+ = \frac{1}{2} [2I_z S_x + i 2I_z S_y] \rightarrow \frac{1}{2} I^-$$

is possible, that both cartesian components can bet ransfered to the observable signal.

Coherence order selective transfer (sensitivity enhancement)



$$\sigma(a) = \frac{1}{2} \left[ -i \, \mathbf{2} \mathbf{I}_z \mathbf{S}_x - \mathbf{2} \mathbf{I}_z \mathbf{S}_y \right] \exp \left\{ -i \left( \Omega_S t_1 - \phi_1(z) \right) + \frac{1}{2} \left[ i \, \mathbf{2} \mathbf{I}_z \mathbf{S}_x - \mathbf{2} \mathbf{I}_z \mathbf{S}_y \right] \exp \left\{ i \left( \Omega_S t_1 - \phi_1(z) \right) \right\}$$

Transformation of both cartesian components (relevant interactions only):

$$2I_{z}S_{y} \xrightarrow{\frac{\pi}{2}(I_{x}+S_{x})} -2I_{y}S_{z} \xrightarrow{\delta\pi J 2I_{z}S_{z}} I_{x} \xrightarrow{\frac{\pi}{2}I_{y}} -I_{z} \xrightarrow{\pi I_{x}} I_{z} \xrightarrow{\frac{\pi}{2}I_{x}} -I_{y} \xrightarrow{\pi I_{x}} I_{y}$$

$$2I_{z}S_{x} \xrightarrow{\frac{\pi}{2}(I_{x}+S_{x})} 2I_{y}S_{x} \xrightarrow{\pi I_{x}} 2I_{y}S_{x} \xrightarrow{\frac{\pi}{2}(I_{y}+S_{y})} -2I_{y}S_{z} \xrightarrow{\Delta\pi J 2I_{z}S_{z}} I_{x}$$

the overall transfer with these transformation is given by:

$$\frac{1}{2}[-i2I_{z}S_{x}-2I_{z}S_{y}] \longrightarrow \frac{1}{2}[(-i)I_{x}-I_{y}] = \frac{(-i)}{2}I^{-} + \frac{1}{2}[i2I_{z}S_{x}-2I_{z}S_{y}] + \frac{1}{2}[iI_{x}-I_{y}] = \frac{i}{2}I^{+}$$

with relative gradient strengths  $\gamma_S G_1 = \gamma_I G_2$  the transfer  $I_z S^+ \to \frac{1}{2} I^-$  is selected. This results in the following observed signal:  $Signal_N(t_1, t_2) = \frac{1}{2} \exp\{-i\Omega_S t_1\} \exp\{i\Omega_I t_2\}$ 

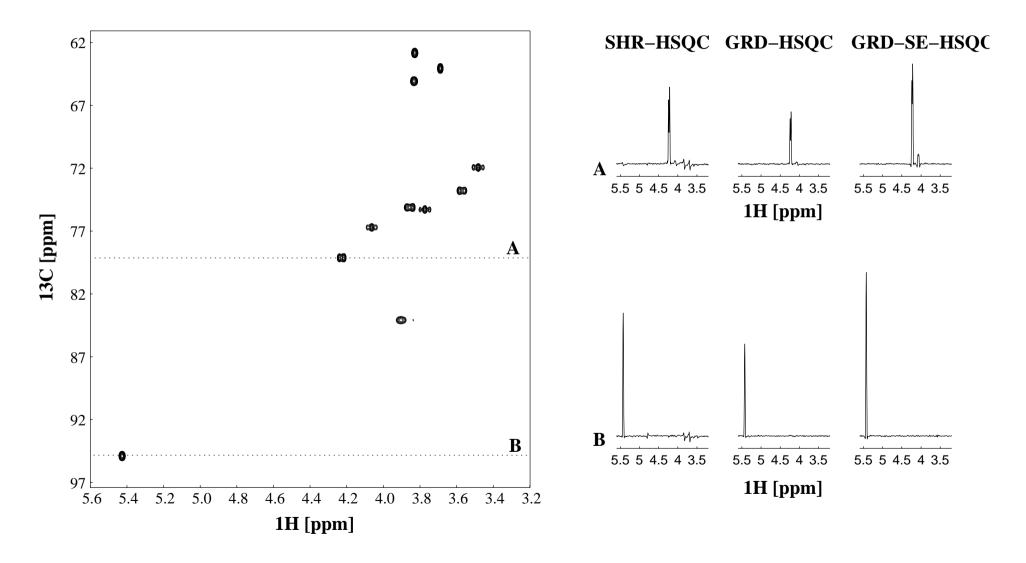
With the gradient strengths  $\gamma_S G_1 = -\gamma_I G_2$  and  $\phi_1 = -1$  the transfer  $I_z S^- \to \frac{1}{2} I^-$  is selected. This results in the following observed signal:  $Signal_P(t_1, t_2) = \frac{1}{2} \exp\{i\Omega_S t_1\} \exp\{i\Omega_I t_2\}$ 

The echo-antiecho transformation gives the final twodimensional phase sensitive signal.

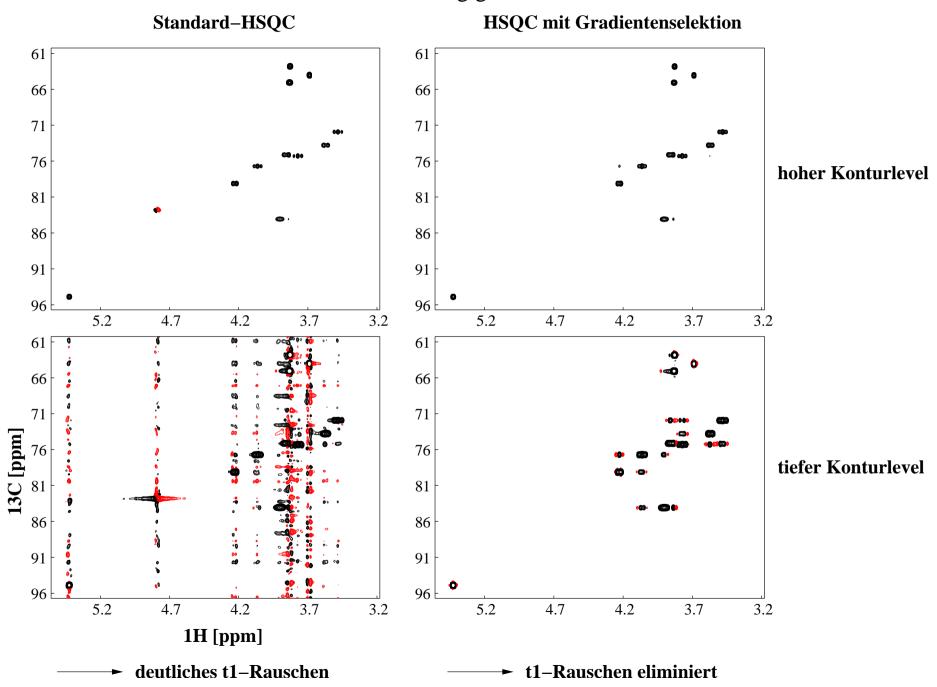
This signal intensity is 2 times more intensive compared to the previous selection by gradients, if the additional pulses (imperfections!) and time intervals (relaxation!) cause no further signal loss.

#### Comparison of different techniques: <sup>1</sup>H, <sup>13</sup>C HSQC on sucrose

(ca. 15 mM sucrose in  $D_2O$ , 400 MHz, 128\* (F1) 1024\* (F2) data points, NS = 16, total duration ca 1.5 each spectrum)



#### Artifact elimination using gradient selection



#### Origins of the $t_1$ -noise

In the chosen pulse program for the standard HSQC the rf phases allow that the resonances from protons not bound to a <sup>13</sup>C-spin (natural abundance <sup>13</sup>C ca. 99% !!) result in observable signal in the FID.

These must be suppressed in a difference manner by a proper phase cycling!

Instabilities of the spectrometer causes a not perfect reproducible signal in different scans, therefore along  $t_1$  a variation (noise) of the not perfectly suppressed signal results

In the gradient experiment the signals from protons not bound to  ${}^{13}$ C-spins are suppressed effective in each single scan. This minimizes the residual  $t_1$ -noise to a large extend.

#### Water suppression in protein NMR spectroscopy

16-bit analog to digital converter: intensity is measured in integer numbers ranging from  $-2^8$  bis  $+2^8-1$ ,

that is from -32768 bis +32767

proton concentration of  $H_2O$  2 x 55 mol/l,  $1.1 * 10^2 M$ 

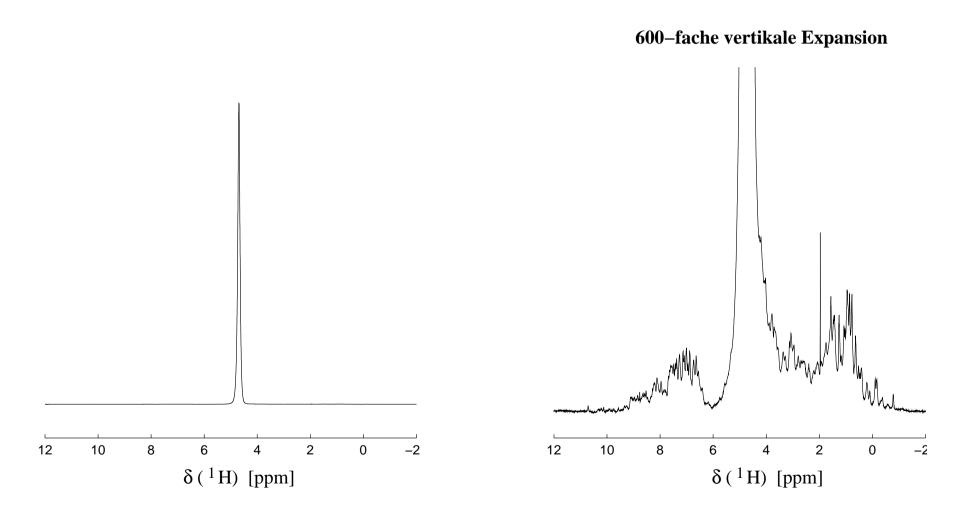
typical protein concentration 1 - 2 mM

→ H<sub>2</sub>O signal is ca. 50000 - 100000 times more intense than an individual protein proton.

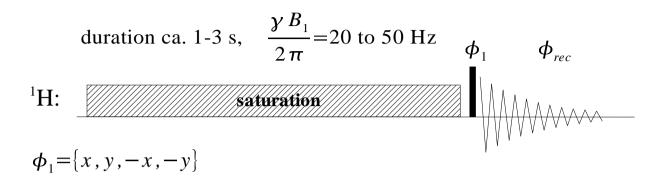
It is often not possible to detect protein signals near the not suppressed water!

The essential detection of the amide protons requires samples in  $H_2O$ , because in  $D_2O$  the exchange of labile protons with deuterons results in their signal loss.

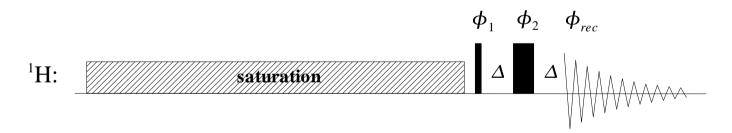
## Onedimensional <sup>1</sup>H NMR experiment on protein with out water suppression (2 mM lysozyme in 90% $H_2O / 10\% D_2O$ , 400 MHz, NS = 32)



#### Onedimensional <sup>1</sup>H NMR experiments with presaturation for water suppression



additional spin echo



$$\Delta$$
 ca. 100  $\mu$ s
$$\phi_1 = \{4x, 4y, 4(-x), 4(-y)\}$$

$$\phi_2 = \{x, y, -x, -y\}$$

$$\phi_{rec} = \{x, -x, x, -x, y, -y, y, -y, -x, -x, -x, -x, -y, y, -y, y\}$$

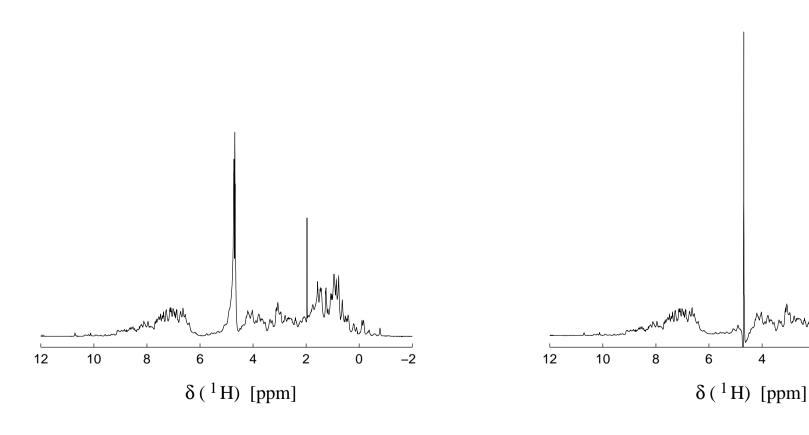
## Onedimensional <sup>1</sup>H NMR experimente on a protein in H<sub>2</sub>O in water using presaturation for water suppression

(2 mM lysozyme in 90% H<sub>2</sub>O / 10% D<sub>2</sub>O, 400 MHz, NS = 32, saturation 3 s,  $\frac{\gamma B_1}{2\pi}$  = 49 Hz)

#### Vorsättigung

#### Vorsättigung mit Hahn Echo

2

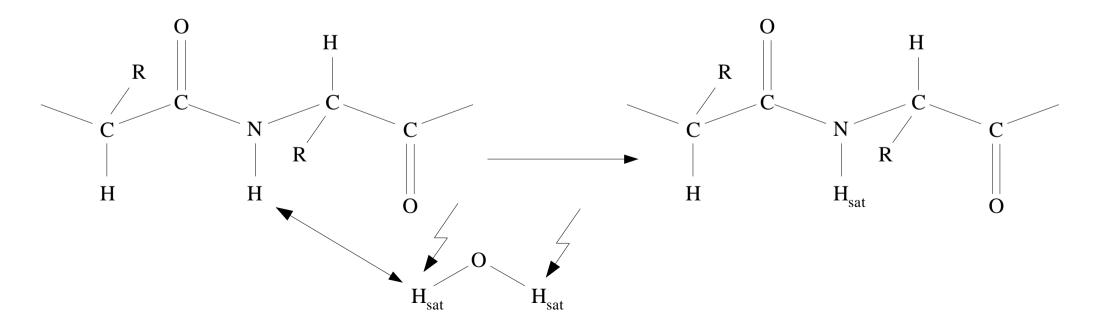


#### Problems with water suppression by presaturation

Saturation is frequency selective, protein resonances near the water are also saturated

Residual water signal is often most intense signal in the spectrum even after presaturation. In 2D experiments this results in intense  $t_1$ -noise at  $F_2 = d(H_2O)$  and causes baseline distortions.

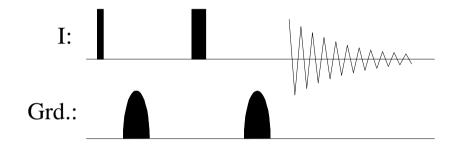
The exchange of labile protons (e.g. amide protons) with water results in saturation transfer, if the exchange rate is faster than the longitudinal relaxation of the exchanging proton



#### Water suppression with pulsed field gradients

Basic principle: spinecho with two gradients of equal strength and duration

→ good selection of refocusing in a single scan



Goal: The  $180^{\circ}$  pulse shall only refocuse protein magnetization but not the water spins (H<sub>2</sub>O antiselective pulse) in this case the difference of the gradients (=0) acts on the protein signals, and water protons are affected by the sum of the two gradients

right efficient dephasing of the water magnetization by the gradients

Watergate

$$180^{\circ}(x)$$

I: 90°(-x)<sub>H2O</sub> 90°(-x)<sub>H2O</sub>

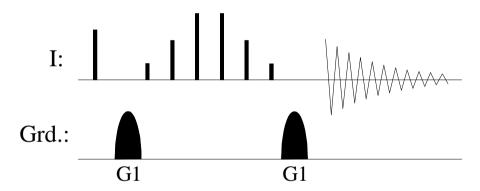
Grd.: G1 G1

Rotation anglel for  $H_2O$  protons:  $-90^{\circ} + 180^{\circ} - 90^{\circ} = 0^{\circ}$ 

Rotation angle for protein protons: 180°

Binomial (3-9-19) watergate

G1



$$3a(x) - t - 9a(x) - t - 19a(x) - t - 19a(-x) - t - 9a(-x) - t - 3a(-x)$$
  
 $26a = 180^{\circ}$ ,

excitation maximum at ca. 1 / (2t) relative to carrier frequency

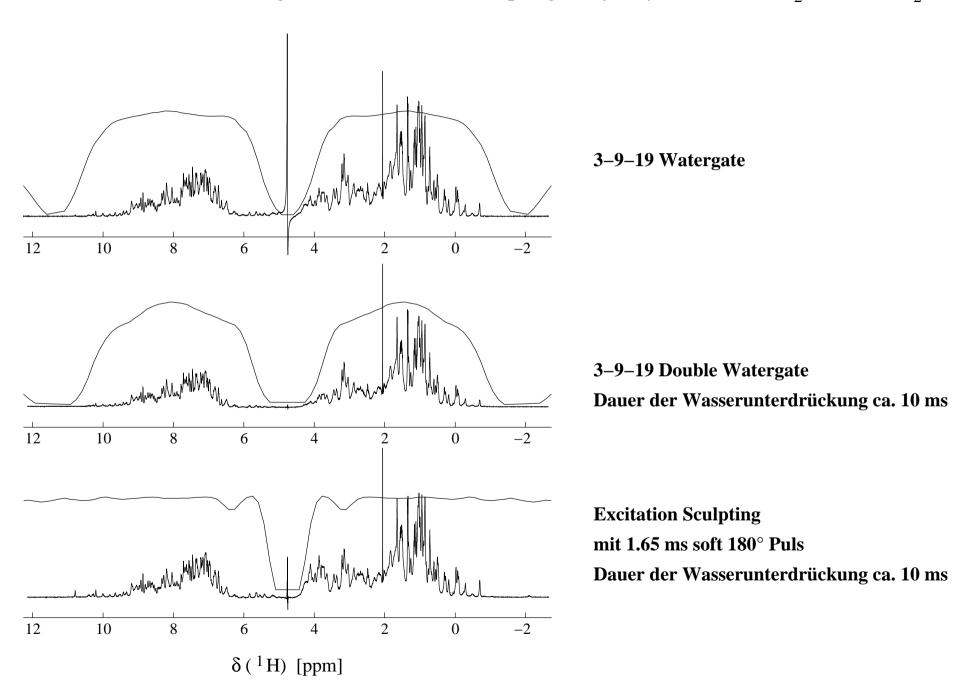
double watergate (excitation sculpting)

G2

G1 G2

Double watergate improves water suppression and compensates the off-resonance effects caused by the first selective pulse.

Binomial (3-9-19) watergate and excitation sculpting on lysozyme in 90 % H<sub>2</sub>O / 10 % D<sub>2</sub>O



#### Additional features of watergate-techniques

No continuous saturation during the relaxation delay between the scans

reduces saturation and saturation transfer

Water suppression is frequency selective, protons near the water resonance are also affected

The water suppression occurs after the exciting pulse. During the time period necessary for water suppression transverse relaxation reduces the initial signal intensity.

Example: Excitation sculpting, selective pulse 2.5 msm, gradient puls ca. 0.5 ms

→ total duration ca. 12 ms

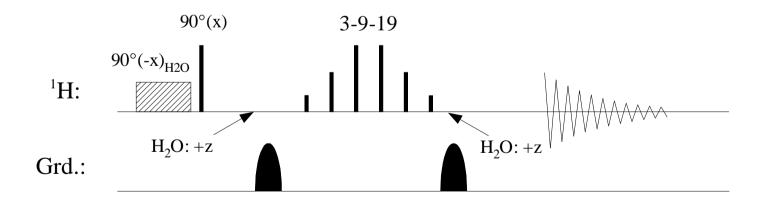
 $T_2(H^N, lysozyme)$  ca. 35 ms signal loss of ca. 30%!

#### Minimal water saturation - Water-flip-back

Signal accumulation faster of the logitudinal relaxation of water results in a saturation transfer of saturated water protons to fast exchanging amide protons.

This results in a reduced signal for fast exchanging protons.

If the applied pulses rotate the water magnetization to +z (equilibrium) at the beginning of the FID, the saturation transfer will be minimized.



### Aeqous samples and radiation damping <sup>1</sup>H pulse length calibration

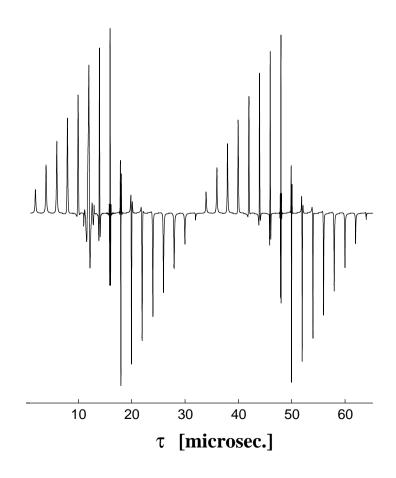
$$I_z \xrightarrow{\omega_1 \tau I_y} I_z \cos(\omega_1 \tau) + I_x \sin(\omega_1 \tau)$$

#### 1% H2O in D2O (400 MHz)

# 20 40 60 80

 $\tau$  [microsec.]

#### 90% H2O / 10% D2O) (700 MHz)

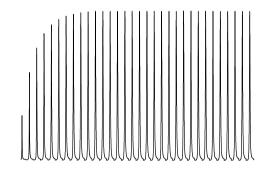


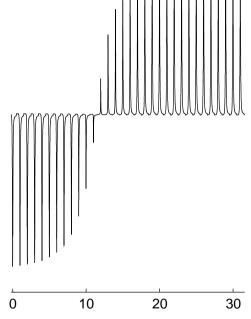
#### Determination of radiation damping time constant at 700 MHz

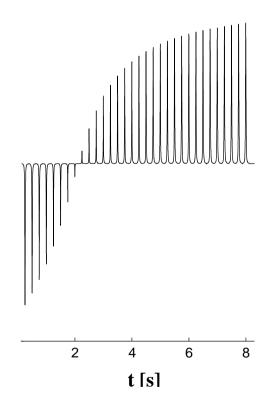
$$90^{\circ} - t - Grad. - 90^{\circ} - acq.$$

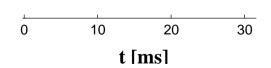
$$180^{\circ} - t - Grad. - 90^{\circ} - acq.$$

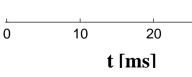
$$90^{\circ} - t - Grad. - 90^{\circ} - acq.$$
  $180^{\circ} - t - Grad. - 90^{\circ} - acq.$   $180^{\circ} - t \text{ (weak Grad.)} - 90^{\circ} - acq.$ 



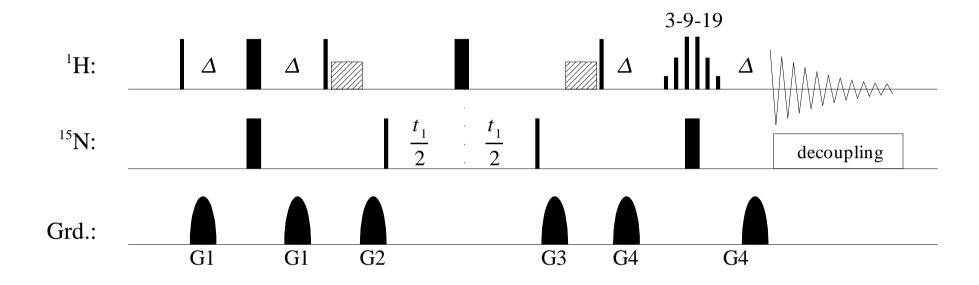








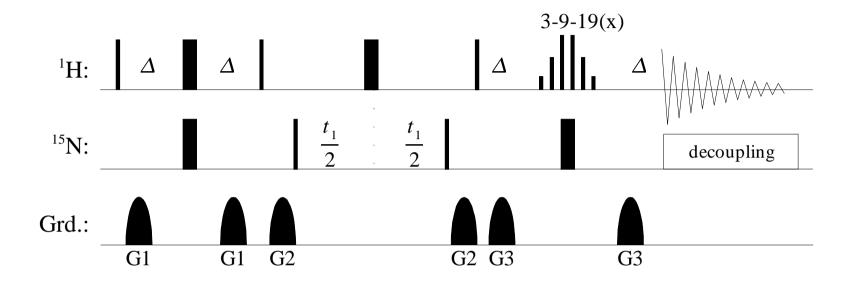
#### Water-flipback in a <sup>1</sup>H, <sup>15</sup>N HSQC with selective pulses



Gradient pairs 
$$G1 - 180^{\circ}$$
 (H) - G1 and  $G3 - 180^{\circ}$  (H) - G3

suppress radiation damping by dephasing anf rephasing of the water magnetization during the INEPT steps selective water flipback pulse : 2 ms rectangular pulse or shaped pulse

#### Water-flipback in a <sup>1</sup>H, <sup>15</sup>N HSQC without selective pulses (FHSQC, fast HSQC)



Gradient pairs

G1 - 180° (H) - G1 and

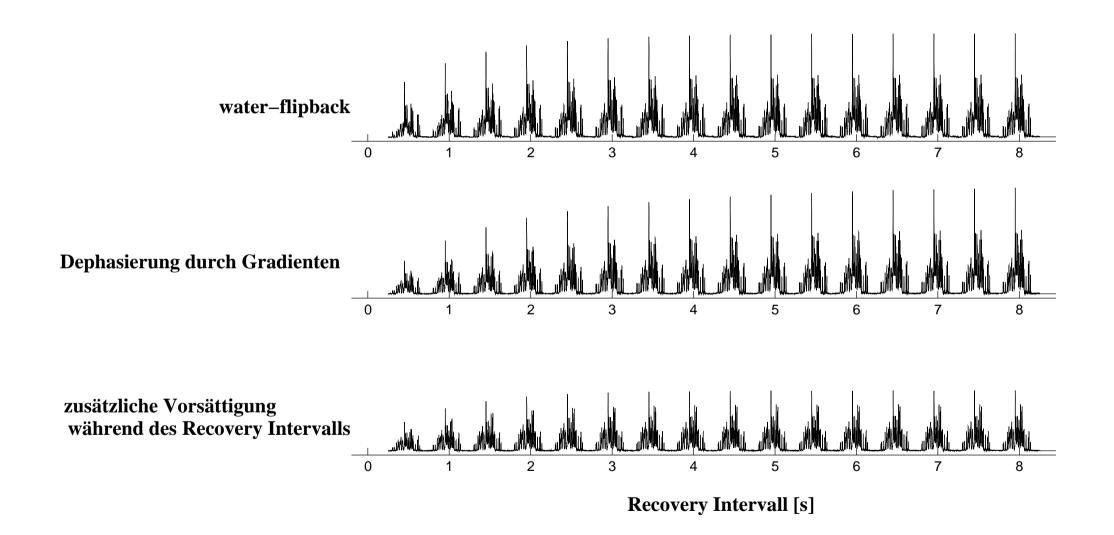
 $G2 - 180^{\circ} (H) - G2$ 

suppress radiation damping by dephasing – rephasing of the water during the INEPT steps and during <sup>15</sup>N evolution

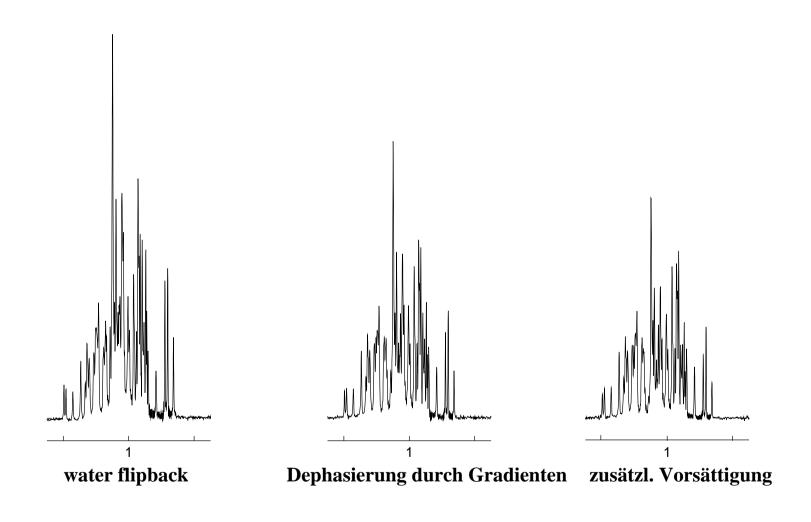
<sup>1</sup>H pulse phases are chosen that H<sub>2</sub>O magetization is along +z (quilibrium) at the beginning of the FID

binomial 3-9-19 watergate suppresses residual transverse water magnetization

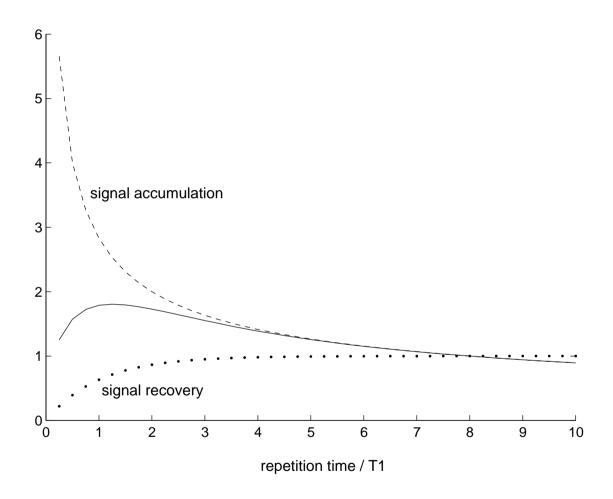
#### Different water suppression schemes in 1D <sup>1</sup>H, <sup>15</sup>N HSQC experiments



Different water suppression schemes in a 1D <sup>1</sup>H, <sup>15</sup>N HSQC with a relaxation delay of 1 sec

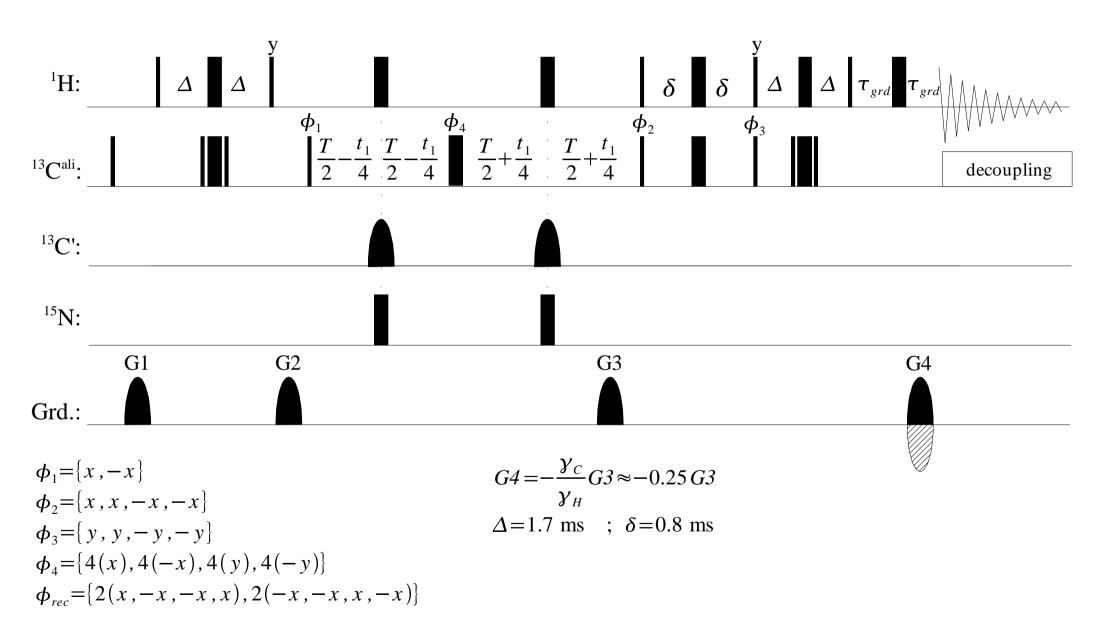


Sensitivity of a NMR experiment as function of the repetition rate



-> optimal interscan delay approx. 1.25\*T1(spin diffusion averages proton T1 in macromolecules)

<sup>1</sup>H, <sup>13</sup>C constant time HSQC for <sup>13</sup>C, <sup>15</sup>N labeled proteins



Recording of the antiecho: inversion of G4 and  $\phi_3$ 

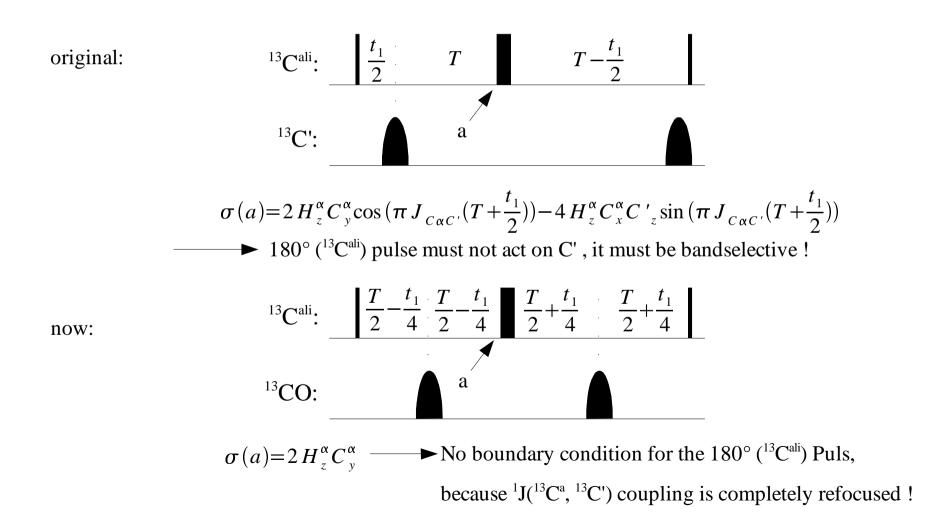
#### Remarks to the presented pulse sequence

Before the initial INEPT the <sup>13</sup>C equilibrium z-magnetization is dephased by 90°(<sup>13</sup>C) - gradient dephasiert.

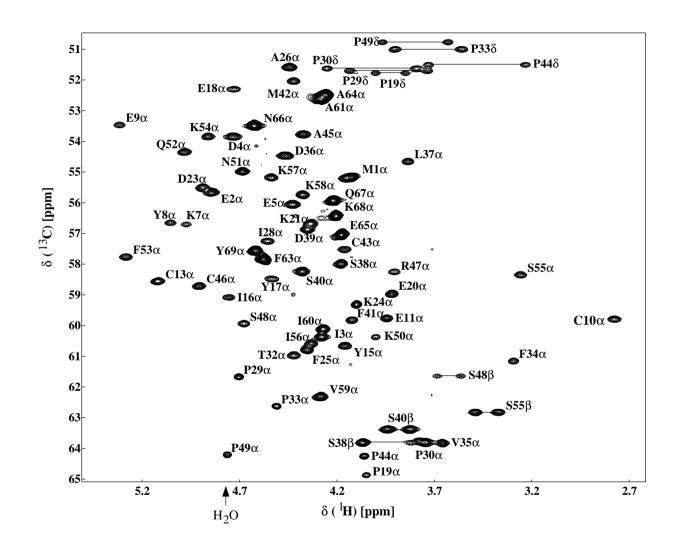
This ensures that no observable magnetization results from this equilibrium magnetization.

The <sup>1</sup>J(<sup>13</sup>C<sup>a</sup>, <sup>13</sup>C') coupling during the evolution period is refocused by selective 180°(CO) pulses.

In the original ct-HSQC these 180° pulses are applied at different time points:



Water suppression in a  $^{1}$ H,  $^{13}$ C ct-HSQC with gradient selection (ca. 2 mM  $^{13}$ C,  $^{15}$ N *G. theta* Rubredoxin (70 aa) in H<sub>2</sub>O /D<sub>2</sub>O 9:1, 600 MHz)



Intensity of residual  $t_1$ -noise at  $F2(H_2O)$ : only 3-5 times the thermal noise level !!